

The importance of the cashew nut (*Anacardium occidentale* L.) coat: a review

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ABSTRACT

The cashew (*Anacardium occidentale* L.) is a major source of income for farmers in the Northeast of Brazil. The cashew nut is composed of three parts: shell, nut, and brown film- known as coat. The coat represents 1% to 3% of the nut total weight and is a rich source of polymeric hydrolysable tannins, as polyphenols. The lipid fractions are particularly comprised by fatty acids, oleic (C18: 1) and linoleic (C18: 2). This study reviewed the cashew nut coat, in light of the scientific advances achieved in recent years. The aspects examined were: separation process between the coat and the cashew nut, coat chemical composition considering nutrients and other bioactive compounds of primary and secondary metabolism, biological and microbiological activities and technological applications.

Keywords: Cashew nut coat, phenolic compounds, *Anacardium occidentale* L.

INTRODUCTION:

The cashew tree (*Anacardium occidentale*) is a Brazilian plant widely available in the coastal region, extending from the Amazon to the Northeast. It is distributed in many tropical regions of the world, among which are: Mozambique, Tanzania, Kenya, Guinea Bissau, Indonesia, Thailand, Vietnam [1, 2], and India [3]. The cashew nut trade began in the early 1920s. India was a pioneer in processing and trading these nuts in industrial scale, and it remains the top cashew nut producer in the world, followed by Vietnam and Brazil [4].

In Brazil, the cashew nut agribusiness is concentrated in the Northeast region. Together, the states of Ceará, Piauí, and Rio Grande do Norte are responsible for 95% of the total production. The sale of cashew nut contributes significantly to the local economy, creating more than 300,000 jobs in the Northeast [5]. According to IBGE (the Brazilian Institute of Geography and Statistics), Brazil has close to 775,000 hectares dedicated to cashew groves, area that practically remains unchanged over the past 10 years. The total cashew nut production ranges around 276,000 tons per year [6].

The cashew tree has been considered a great source of phytotherapies for centuries. Its application in popular medicine is described in the literature. Common uses include: analgesic [7], diuretic [8], antiseptic for oral hygiene [9], asthenia treatment, respiratory problems, genital infections, and skin diseases [10].

Literature reports evidences of its successful uses as a hyperglycemic [11], an antimicrobial [12, 13], antioxidant [14, 15], anti-inflammatory [16, 17], anti-ulcerogenic [18], and antiophidic [19].

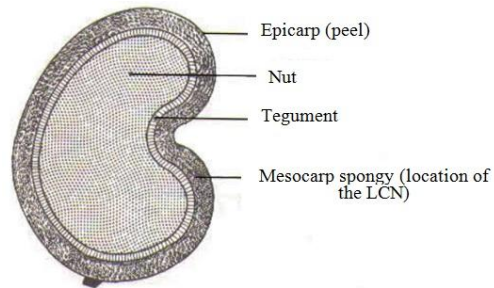
To make the observations, chemical constituents were isolated and identified from many parts of cashew tree.

From the chemical study of the leaf, it is highlighted the occurrence of flavonoids such as: agathisflavone, apigenin, kanferol, myricetin, quercetin, quercetin-3-O-glicopiranosil, quercetin-3-O-ramnopiranosil, robustflavone, and amentoflavone [20]. The Ethyl gallate was isolated from the flowers [21]. Its bole shell shows the gallic acid as a major component. The steroids were obtained as tannins hydrolysis product: myo-inositol, cholesterol, campesterol, stigmasterol, and sitosterol [22]. The *occidentosideo(-)-salipurposideo* were isolated from the nut shells [23, 24], naringenin, naringenin-7-O-(6"-O-p-coumaroyl)- β -Dglycosylates [25], naringenin-5 β -glycosylates[23]. Various phenolic sources were selected for the coat study, in which (+)-catechin and (-)-epicatechins were isolated [21]. In addition, pharmacological activities were tested in one of the bioactive compound classes and those that attracted more interest were phenolic lipids, mainly for its antioxidant properties [26, 27].

Although there are many studies on the cashew tree, a detailed research on the coat is scarce. The subject has received more attention only in recent years, for its functional potential [4]. However, there is little information on the chemical structure and the physicochemical and functional properties of the cashew nut coat. Therefore, in this review, four themes will be addressed: (i) cashew nuts processing and waste generation (coat) (ii) chemical and metabolic composition, (iii) biological activities, and, (iv) possible industrial applications.

• THE CASHEW NUT

The cashew nut consists of three parts: shell, coat and nut. The weight of a nut may vary from 2g to 30g, and the average weight is around 7.0g. The Figure 1 shows a cross section of cashew nuts.

Figure 1. Cross section of a cashew nut

Source: [4].

The shell, which is 65% to 70% of the nut's weight, consists of an epicarp coriaceous, crossed by a spongy mesocarp, whose alveoli are filled with a caustic and flammable liquid, the CNSL (Cashew Nutshell liquid) [28]. The film, or coat, represents around 3% of the nut's weight, it is rich in tannins [29], and the nut, the edible part, formed by two ivory cotyledons, represent approximately 28% to 30% of its weight. In the industrial process, the average yield is only 21% [30]. The CNSL consists of a dark brown liquid, viscous, acre and caustic, rich in phenolic compounds (anacardium acid, cardol, cardonol, and 2-metilcardol) [31, 32].

The cashew nut coat is a thin protective layer of the nut (Figure 1). It was initially utilized to maintain the operation of boilers, on CNSL extraction and in cattle feed [33]. These remain the most used applications to date. Numerous authors have observed the coat's phenolic activity, opening doors for many studies related to tannins applicability. Varnishes manufacture, elaborated from resins obtained from tannin extracts of the coat, are cited as one of its industrial applications [34]. The cashew nut coat represents an environmental problem for the producing regions because due to the high production of nuts and, consequently, coat, the manufacturing plants have no use for this residue, except for utilizing it to power boilers [35].

• CASHEW NUT PROCESSING

The cashew nut processing aims to obtain the whole nut, fully dehulled, white-ivory, spotless, and good sized [36]. The cashew industrialization (peduncle and nut) can be divided, at least, in two types of processing: the nut processing industry and the peduncle manufacturing industry. The first aims to obtain cashew nuts (CN), with cashew nutshell liquid (CNSL), nut shell, film (rich in tannin), and cashew nut oil as its main by-products. The peduncle manufacturing industry, in turn, has segments in beverages, sweets, condiments, flours, and feeds industries, among others.

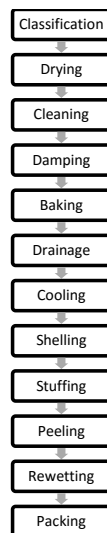
The cashew nut processing is performed, predominantly, by mechanized and semi-mechanized systems in large factories. This process may also take place in establishments classified as micro or small homemade production, cooperatives, and producer associations (with capacity of process up to one ton/day). The average yield of these processes is approximately 23%, i.e., 4.35 kg of raw nuts are necessary to produce one kilogram of nut [28].

The handmade process of nut processing is still widely adopted in small farms in the Northeast of Brazil, especially in the states of Piauí and Bahia. Other countries (Africa, India, Sri Lanka, and Vietnam) also utilize this process due to the availability of low cost labor and for its higher yield of the whole nuts, around 85% to 95% [37]. This process has many drawbacks, linked mainly to poor hygiene conditions of the settings [38].

The Brazilian industry of cashew nut processing utilizes both mechanized and semi-mechanized segment. The mechanized segment consists of 12 processing plants with the capacity of processing about 90% of the Brazilian production [39], and the semi-mechanized segment consists of over one hundred mini factories, with capacity of processing 20,000 tons per year (Paiva, S. N. et al., 2006). The mechanized industrial processing of cashew nuts is represented according to the Flowchart 1.

• NUT AND COAT SEPARATION PROCESSES FROM CASHEW NUT

Flowchart 1: Cashew processing phases in mechanized system



The processing starts with the nuts' arrival in the industry. First, they are weighed and taken to sunlight exposure aiming to reduce moisture (from 7% to 10%) to avoid deterioration problems and, thus, leading to the maturation of the nut by the action of infrared and ultraviolet rays [40]. The nuts remain drying for a period that can reach up to seven days, and a classification by size is, then, performed, following a previous cleaning using vibrating screens or perforated plates [41].

The nuts, after being dried, cleaned, and sorted, may be stored in sacks, stacked on pallets arranged on a waterproof floor [42, 43]. Then, they are cleaned and stored in bins, which are immersed in water for a period ranging from 140 to 330 min [44]. Nut roasting is usually carried out in an autoclave at 110°C/10 min, or in a household pan [45].

The CNSL, first noble product of the processing, is extracted during roasting process. The dehulling or shelling may be performed by centrifugation ("Stutervant" process) or cutting ("Oltremare" process) [29].

After shelling, drying aims to reduce nut moisture from 2.5% to 4.0%, wherefore the coat, firmly adhered, becomes brittle and can easily be removed. Drying is performed by greenhouse (between 60°C to 80°C) for 6 to 8 hours. In many cases, the nut is submitted to a wetting process by saturated steam (between 1 or 2 minutes), which facilitates the coat separation from the nut [46].

The nut tends to decrease in size and the coat becomes brittle with dehydration, weakly adhering to the nut. Then, the shelling is carried out taking advantage of this physical change. The coat removal is done by injecting compressed air to break or release the coat, or by a shelling cylinder with brushes, or using an electric rotary cylinder [46, 47].

The shelling with rotary cylinder, activated by an electric motor at low speed, consists of submitting the nuts to friction on a perforated screen, promoting the partial release of the coat. In a shelling cylinder with brushes, the nuts are placed on a wood table or galvanized sheet provided with metal screen, where they are submitted to friction through the bristle brushes to obtain the nuts partially without coat. In any of these operations one may obtain nuts up to 70% without coat. The remaining shell is submitted to a manual scraping process with shelling knives [43].

• COAT CHEMICAL COMPOSITION

Thousands of chemical constituents may be present in plant tissues, although, largely, in low concentrations. Even under these conditions, several of these constituents are responsible for features such as color, flavor and taste, besides nutritional and nutraceuticals effects [48-50].

Among the chemical constituents identified in the cashew nut coat, one can highlight the following classes: terpenes, flavonoids, terpenoids, catechins, epicatechins, tannins, and sterols. Phenolic compounds are considered as the primary responsible for pharmacological activity [51-59].

Studies performed [60] to determine the cashew nut coat (CNC) mineral composition, show that it contains more protein, fiber, calcium, magnesium, phosphorus than corn, but with less energy if compared to values reported for that grain (Table 1).

Table 1: Chemical and mineral composition of the cashew nut coat

ANALYSIS	NTC (g/Kg)	Corn (g/Kg)
Dry matter (DM)	905	890
Crude protein	190	88
Crude fiber	103	22
Ether extract	20,1	38
Ash	20,2	13
Calcium	5,6	0,2
Phosphorus	1,9	2,8
Magnesium	5,8	1,2
Tannins	1,8	--
Metabolizable energy	7,12	14,02

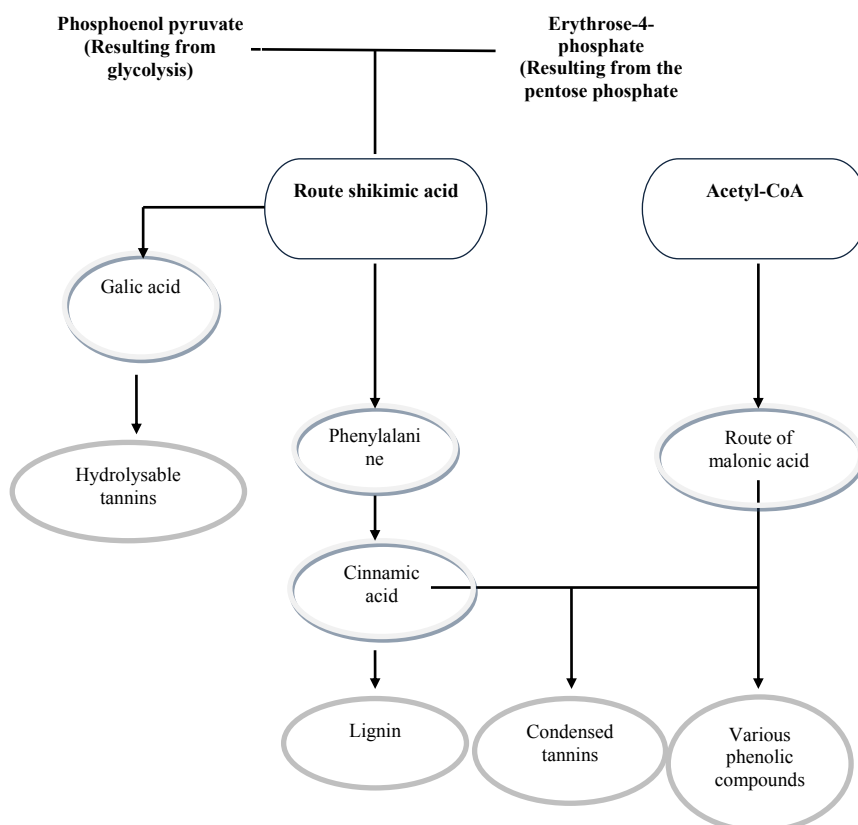
Source: [59].

• METABOLISM, PRIMARY AND SECONDARY METABOLITES

Plants primary biosynthetic process is the photosynthesis, which uses solar energy for organic compounds production, grouped according to common features of primary and secondary metabolites. The primary metabolites, essential for organism survival are: sugars, amino acids, fatty acids,

nucleotides, and polymers. Some of these metabolites are used as precursors in the synthesis of other compounds, in enzymatically catalyzed reactions, such as shikimic acid (precursor of several aromatic compounds), acetate (a precursor of fatty acids, polyphenols, isoprene, prostaglandins, among others), and aliphatic amino acids (alkaloids biosynthesis), summarized in flowchart 2 [61].

Flowchart 2: Some primary metabolites routes



Source: Adpt [62].

• PRIMARY METABOLITES DERIVED OF FATTY ACID PRESENTED IN COAT (CNC)

The fatty acid is an essential primary metabolite for plants and animals. It is derived from a biosynthetic route of acylpolimalonato (AcetylCoA), creating long carbon chains. Its dehydrogenation and/or oxidation may give rise to heterocyclic, triglyceride, and lipid compounds [63].

The cashew nut coat has a high amount of ether extract (Table 1), characteristic that arises one's interest in understanding its lipid composition [64]. The main fatty acids found in the coat are shown in Table 2.

The coat's lipid composition values vary based on different factors such as sample preparation, sample homogenization with solvents, phase separation, and solvents removal. Sample preparation depends on its nature, harvest local, extraction forms, storage period, and temperature [65]. During the lipidic extraction, some conditions are

considered, such as operational costs, implementation speed, ease execution, to include the care to prevent lipid and hydrolysis oxidation and degradation during analysis and storage stages [66].

Table 2 analysis allows the comparison of two extractive methods performed by [67] and [68], with extraction solvents (chloroform/ methanol/ water) and (ethyl acetate/ hexane), respectively.

Table 2: Fatty acid composition CNC

FATTY ACIDS	[67](%)	[68] (g/kg)
Lauric acid (C12: 0)	0,2	-
Myristic acid (C14: 0)	0,3	-
Physeteric acid (C14: 1)	0,4	-
Palmitic acid (C16: 0)	16,4	-
Palmitoleic acid (C16: 1)	1,1	-
Hexadecodienóico acid (C16: 2)	1,4	-
Stearic acid (C18: 0)	6,4	40,9± 6,3
Oleic acid (C18: 1)	35,3	214±33,2
Linoleic acid (C18: 2)	30,4	68,6± 10
Linolenic acid (C18: 3)	5,8	-
Gadoleic acid (C20: 1)	1,6	-
Fatty acids	0,8	-

(-) not analysed.

One of the advantages of the method used by [67] is the formation of a biphasic system from solvents proportions added during the extraction process, based on the liquid-liquid balance theory of three components (chloroform/ methanol/ water). The methodology used by [64] was performed following the method used by [69], which analyzes stearic, oleic, and linoleic acids, for being of great importance in the extract and oil characterizations. Unlike the previous method [67], the authors sought solvents like ethyl acetate and hexane, to replace the chloroform. The analysis based on gas chromatography was coupled to mass spectrum which allowed the quantification of the fatty acids present in the coat.

The stearic acid content (4.1g/100g) in the coat was higher than those found in peanut (1.30g/100g), hazelnut (0.94g/100g), coconut (1.10g/100g), nut (0.55g/100g), pistachio (0.98g/100g), and walnut (1.37g/100g)[70]. The oleic acid content (21.40g/100g) showed a higher concentration than the nut (0.98g/100g), coconut (2.10g/100g), Brazil nut (18.5g/100g), and walnut (11.4g/100g), while the linoleic acid content of the coat (6.9g/100g) was greater than the one present in coconut (0.68g/100g) and macadamia (1.74g/100g) [71-73]. The linoleic acid is a phospholipid component, responsible for

cytoplasmic membrane structure and function [74]. According to that study, a diet rich in polyunsaturated fatty acids has anti-spreader effects of diseases, such as: diabete type 2, hyperlipidemic, cardiovascular disease, inflammatory rise, cancer, and osteoporosis.

• SECONDARY METABOLITES PRESENTED IN THE COAT (CNC) OF *ANACARDIUM OCCIDENTALE* L.

Secondary metabolites are responsible for maintaining the plant, i.e., the interaction with the native environment. Generally, they have a complex structure, low molecular weight, and remarkable biological activity, protecting the plant against pathogens and microbial attacks, and herbivorous actions. It is produced by plants in various stress situations including nutrient deficiency, salinization, water scarcity, and high exposure to UV radiation, leading to the production of more polar compounds [63, 75, 76]. Secondary metabolites are divided into three classes: phenolic compounds (phenols, phenolic acids, coumarins, flavonoids, tannins, and lignins); terpenes (carotenes, steroids, polysoprenos, saponins, and triterpenes), and alkaloids (nitrogenous compounds) [76, 77].

Phytochemical research is one of the most widely used methods for the isolation and purification of secondary metabolites, aiming at identifying the chemical constituents of a given plant specie or evaluating its presence [78].

Whenever there is no chemical study available on species of interest, the preliminar phytochemical analysis may indicate the relevant groups of secondary metabolites therein. The extracts prepared from plants are submitted to tests which allows the characterization of the main groups containing the organic substances of interest [63].

Researchers [79] and [80] identified in cashew nuts classes of secondary metabolites as alkaloids, polyphenols, and saponins. Among the classes of secondary metabolites, the more prevalent in the coat are: tannins, phenolics, and flavonoids (catechin, epicatechin, and epigallocatechin). Studies with coat (CNC) pointed that phenolic constituents highlight the hydrolysed tannins with polymeric proto anthocyanidins. Table 3 presents a phytochemical analysis of the cashew nut coat extracted by solventes of different polarities [68].

Table 3: Coat (CNC) chemical profile of *Anacardium occidentale* L. using different solvents.

PHYTOCHEMICAL	Solvent		
	Ethanol	Ethyl acetate	Acetone
Alkaloids	✘	✘	✘
Carbohydrates	✘	✓	✓
Steroids	✘	✘	✘
Phenolic	✓	✓	✓
Flavonoids	✘	✘	✓
Glycosides	✘	✘	✘
Volatile oils	✓	✓	✓
Triterpenoids	✓	✘	✓
Xantoproteins	✘	✓	✓

(✘) Not detected (✓) Present

Source: [79].

Natural compounds have a different solubility behavior depending on the chemical nature of different functional groups present and may vary from simple to highly polymerized substances. In addition, there is also the possibility of interaction between the various compound classes, such as carbohydrates and proteins. These interactions may form insoluble complexes, thus, compromising the reliability of the obtained data. The development of a single extraction procedure, capable of recovering all compounds present in the sample, is improbable, since a small fraction of these compounds are soluble in the solvent utilized [81-83]. The use of different solvents explains the differences in results shown in Table 3. The CNC ethanolic extracts resulted in the presence of various phytochemical compounds such as triterpenoid,

phenolic, and volatile oils. The extract with ethyl acetate showed a different combination of phytochemicals- phenols, volatile oils, xantho proteins, and carbohydrates. However, the extract containing acetone was effective in dissolving triterpenoids, phenolics, volatile oils, flavonoids, xantho proteins, and carbohydrates [79].

• PHENOLIC COMPOUNDS

The outer layers of plant materials such as shells and coat have a high content of phenolic compounds. They act in the defense against pathogens, parasites and predators' attacks, besides contributing to the formation of a variety of colors found in plants [83].

Studies have shown that the content assessment of phenolic compounds present in the CNC depends on the extraction method and type. In ethanol extracts [4] obtained directly from the coat, the concentration of total phenols is approximately 185.44mg/ Gallic Acid Equivalent (GAE). But [27] used ethanol to prepare the extract, under stirring, at 37°C, for a period of 3hrs and, when analyzing the amount of phenolic compounds, they observed that the coat presented a higher total phenolic content (243mg/GAE) than the ethanolic extract at room temperature (Table 4).

Studies performed with CNC degreasing ethanolic extracts, carried out by [84] showed a different behavior with temperature variation, indicating that elevating temperatures resulted in an increase of the phenol content. They obtained 790.9 mg/EAG as a maximum point (130°C), whereas in low temperature (70°C) there was a decrease in the phenolic compound content (701.2mg/ GAE). The explanation for this variation is a consequence of the conjugates phenolic

compounds released during the thermic treatment and the production of Maillard Reaction Products (MRP) [85-87].

Another related study, [88], analyzed the content yields of soluble and conjugated phenolic compounds in ethanolic extracts of raw coat, when submitted to high and low temperatures. The highest yields of phenolic extracts were $44.2 \pm 1.4\text{g}/100\text{g}$ of degreased flour, and its total phenolic content was $347.99 \pm 6.88\text{g}/\text{g}$ of degreased flour, in which the CNC was submitted to 130°C for 33 min (Table 4).

These results are similar to the study performed by [89] and [90] in hazelnut coat and its derivatives, where roasting (175°C/5 min) increased by 40% the total peanut coat phenolic compounds in comparison to the raw coat. The authors concluded that the thermic treatment increased the phenolic compound concentrations in the coat, and its results were consistent with similar studies performed using hazelnuts and peanuts [90, 91].

Table 4: Content of CNC total phenolic compounds submitted to different methods

Extract	Submitted treatment	Total phenolic compounds (TPC)	Reference
Ethanol	Shaker 37°C/3hrs	243 mg /g GAE	[27]
Ethanol	Ambient temperature	185,44±12,04 mg /g GAE	[4]
Ethanol (80%)	Degreased (hexane)	656,2±23,0 mg /g GAE	[84]
Ethanol (80%)	Degreased (hexane) 70°C/ 6hrs	701, 2 ± 21,1 mg /g GAE	[84]
Ethanol (80%)	Degreased (hexane) 130°C/33min	790,9±15,4 mg /g GAE	[84]
Ethanol (80%)	Degreased (hexane)	269,05±9,77 mg GAE/g _{degreased sample}	[88]
Ethanol (80%)	Degreased (hexane) 70°C/ 6hrs	308,50,9±6,88 mg GAE/g _{degreased sample}	[88]
Ethanol (80%)	Degreased (hexane) 130°C/33min	347,51±9,35 mg GAE/g _{degreased sample}	[88]

Phenolic compounds are the most important products of a plant secondary metabolism, acting on the growth, reproduction and natural development of plants and vegetables. Most of them come from phenylalanine amino

acids and, in some plants, from tyrosine. Although animal tissues do not synthesize phenolic compounds, the presence of this type of structure may be attributed to plants ingestion [92, 93]. The three largest groups of phenolic compounds

found in animal diet are flavonoids, phenolic acids, and polyphenols (tannins). Phenolic acids comprise benzoic acid and its derivatives (hydroxybenzoic, gallic, and ellagic, among others), cinnamic acid and its derivatives (coumaric, caffeic, ferulic, and chlorogenic, among others). The tannins are polymers with a high molecular weight, divided into two classes: hydrolysable tannins- comprising gallic acid polymers or ellagic (found in fruits and nuts), and the condensed tannins, catechin, or epicatechin polymers [61].

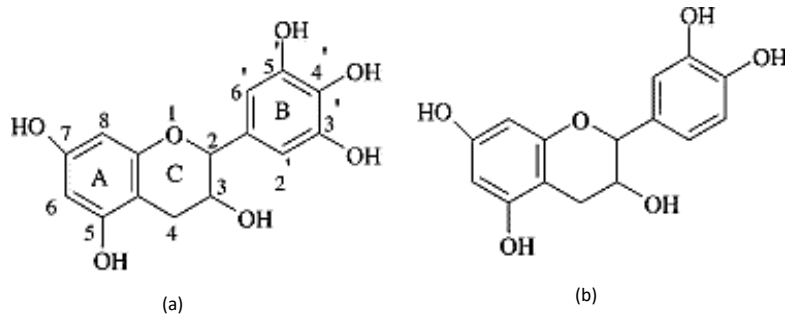
Several phenolic compounds have been found in CNC, such as tannins, flavonoids, catechins and epicatechins, and, in smaller proportions, phenolic acids (synergic, gallic, and coumaric). Studies performed [84] highlight that the values of these acids in extreme conditions can reach $0.974 \pm$

0.030 ; 5.705 ± 0.001 ; and 0.693 ± 0.043 mg/g MS degreased (Table 4). These acids are also present in other nuts for instance cashew nuts, hazelnuts, and pine coat [89, 94, 95].

• TANNINS

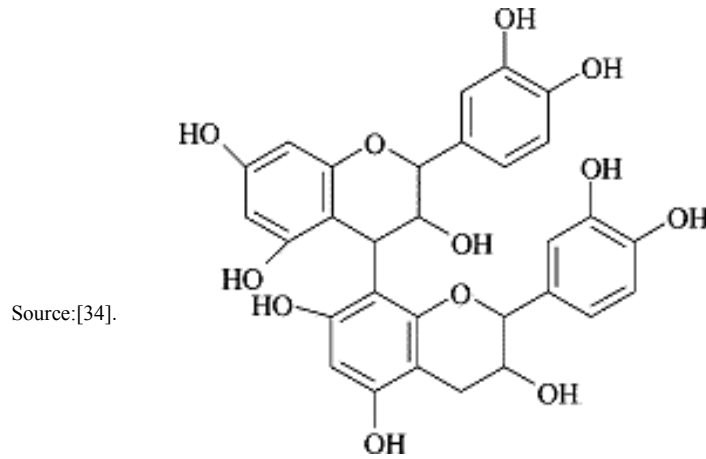
The tannins of cashew nut coat have been studied over the years as a hydrolysable tannins source. The tannin quantitative estimation in the CNC shows that from 82.5% of total polyphenols, 80% are tannins, while the remainder are phenolic constituents [34, 96]. Further studies elucidated and identified the tannins present in the coats (Figures 3 and 4), indicating the presence of procyanidin as its main constituent [34].

Figure 3: Tannins structure of cashew nut coat (a) delphinidin and (b) cyanidin



Source:[34].

Figure 4: Existing catechin in the cashew nut coat.



• CATECHINS

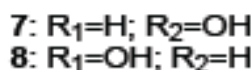
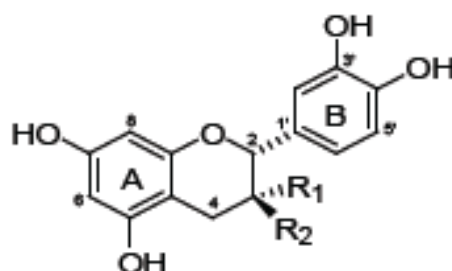
Catechins belong to the flavonoid group of flavanols class. They present a phenyl benzopyran basic skeleton without the carbonyl at carbon 4 and without unsaturation in linking carbons 2 and 3 (Figure 5) [63, 97]. Catechins have beneficial effects on human health, (+) – catechin and (-) - epicatechin have recently received much attention being considered as protective agents against cardiovascular disease and cancer [86, 98, 99].

Researchers [84, 88] observed that the catechin, epicatechin, and epigallocatechin content found in the degressed samples

of cashew nut coat were 47.28mg/g; 28.29mg/g; and 2.0mg/g, respectively.

While evaluating the cashew nut coat extract, one can observe that the HPLC/MS spectrum of the phenolic extract reveals two prominent peaks with maximum absorption at 278nm, highlighting the presence of phenolic compounds such as the catechins[68]. Authors reported the presence of catechins and epicatechins in the coat, through analysis of RMN spectrum of samples isolated from fractioning by column chromatography in silica gel and in ethyl acetate phase (Figure 5)[4].

Figure 5: Structures of (+) - catechin and (-) - epicatechin isolated in CNC



Source: [4].

• OTHER CLASSES OF SECONDARY METABOLITES

The exposure of the human body to free radicals from various sources, leads to the development of defense mechanisms (endogenous defenses) to eliminate these free radicals [100, 101]. These endogenous defenses may be enzymatic or non-enzymatic. The enzymatic antioxidant defenses are in large number and are located throughout the body, both intracellular and extracellular. Among others, the superoxide dismutase, catalase, glutathione peroxidase, and glutathione reductase are examples of these defenses. Non-enzymatic antioxidant defenses highlight compounds such

as glutathione, γ -tocopherol (vitamin E), ascorbic acid (vitamin C), lipoic acid, and carotenoids [101, 102].

Bioactive compounds in cashew nut coats were analysed and it was observed the presence of these compounds in greater quantities than those found in the cashew nut (Table 6) [68]. The presence of high amounts of carotenoids (β -carotene, lutein, and α -zeaxanthin) in the coat is of great importance for the development of cashew nuts during germination. The coat protects it from insects, microbial infection, and sunlight through the enclosure containing the cashew nutshell liquid (CNSL)[64]. After breaking the shell, the antioxidant protection function contained in the coat is

activated [103]. According to [104], carotenoids play an important role in cancer prevention and atherosclerosis.

Table 6: Carotenoids, tocopherols and thiamine in the cashew nuts coat

Bioactive compounds	Concentration (mg/KgDM)
β-carotene	218 \pm 11,8
Lutein	525 \pm 45,2
α-zeaxanthin	7,0 \pm 2,2
α-tocopherol	10,1 \pm 0,7
γ-tocopherol	10,6 \pm 0,6
Thiamine	3,0 \pm 0,5

¹ Values are means \pm standard deviation of 10 separate determinations (n = 10).

Source: [64]

The levels of α - and γ -tocopherols in cashew nut coat are 10.1 and 10.6mg/kg of MS, respectively (Table 6). Tocopherols are reported to constitute an essential part of biological membranes, showing a protective role against lipid peroxidation of membrane, lipoproteins, and fats [105]. Thiamine was also found in CNC (3.0mg/kg of MS).

The coat has a high amount of functional compounds, but its composition may not be described in its entirety because natural constituents are yet to be studied. Moreover, a great number of identified compounds, already isolated and with determined chemical structure, have no specific biological activity, either in terms of their functions, or potential uses, especially therapeutic.

• ANTIOXIDANT ACTIVITY

Research involving antioxidant compounds from natural sources, as plant extracts and its components, have been extensively developed for utilization in foods, in industrial applications, and in animal organisms. This includes different parts, such as seeds (soybeans, peanuts, cotton, mustard, canola, rice, and sesame), fruits (grapes, citrus fruits and peppers), leaves (green tea, rosemary, thyme, and oregano), and others (onion seedlings, sweet potato, and oats) [106-108].

The search for natural antioxidants has been intensified since the 1980s, in an attempt to replace total or partially

synthetic antioxidants. Deleterious effects are attributed to the animal organism when used in high doses [109]. The possible adverse health risks that the irregular and/or indiscriminate use of synthetic antioxidants may present to humans contribute to the high rejection of this additive type. Emphasis has been placed on the identification and purification of new natural compounds with antioxidant activity, which can act alone or synergistically with other additives to prevent oxidative deterioration of foods and restrict the utilization of synthetic antioxidants [89].

Natural antioxidants attract particular interest, especially the ones present in popular foods. Diseases as cancer, atherosclerosis, arthritis, diabetes, and heart-related problems, along with processes responsible for the body's aging may be related to the presence of ROS (reactive oxygen species) in the body [110, 111]. Studies indicate that certain bioactive compounds present naturally in foods may inhibit these processes, due to natural antioxidant qualities [112].

In vitro techniques allow a quick selection of substances and/or potentially interesting mixtures. The principles which guide these methods include the capture of the peroxy radical (ORAC, TRAP), metal reduction power (FRAP, CUPRAC), capture of the hydroxyl radical (deoxyribose method), capture of organic radicals (ABTS, DPPH), quantification of products formed during lipid peroxidation

(TBARS, LDL oxidation, and co-oxidation of β - carotene), RANCIMAT, standard automated method to determine the oxidative stability of oils, fats and lipid food [113-117].

Numerous studies and different methodologies were applied to the cashew nut coat to measure its antioxidant activity, such as DPPH (2,2-diphenyl-1-picryl-hydrazyl), co-oxidation of β -carotene/linoleic acid, ABTS (2,2'-bis-azino 3-ethylbenzthiazoline-6-sulfonic), ORAC (kidnapping of peroxy radical), capture of hydrogen peroxide (H₂O₂), kidnapping of hydroxyl-deoxyribose radical, inhibition of LDL oxidation (inhibition of the lipid peroxidation), FRAP (antioxidant parameter of the reduced ferric ion), and the RANCIMAT [117-122].

The analysis of the cashew nut coat antioxidant activity was performed using different methods. With the ABTS method, the antioxidant activity was measured by the samples

discoloration with ABTS + radical concentration in relation to the coat extract and a synthetic antioxidant, BHA (butylated hydroxyanisole), comparing them. Its potential was also measured using the following methods: Kidnapping of hydrogen peroxide (H₂O₂), kidnapping of hydroxyl-deoxyribose radical, LDL, and FRAP (Table 7)[27].

The results are expressed in EC₅₀ (mg.mL⁻¹), which correspond to the amount of extract required to reduce the DPPH radical in 50%. Thus, the smaller the EC₅₀, the better is the extract antioxidant capacity is. From the data obtained in different tests, it became evident that the CNC effectiveness order is: ABTS> superoxide> deoxyribose> LDL> FRAP (Table 7). Based on the results, one can say that the coat extract is a more potent antioxidant than the ferric reducer.

Table 7: CNC antioxidant activity

Antioxidant activity method	EC ₅₀ (µg/mL)
ABTS	1,30±0,02
FRAP	6000±0,24
LDL	24,66±0,32
Abduction of hydrogen peroxide (H ₂ O ₂)	10,69±1,13
Abduction of hydroxyl-radical deoxyribose	17,70±0,05

Source: [27]

Other authors [123] investigated the nut coat ethanolic extract potential to kidnap the DPPH radical. The results obtained, expressed by antioxidant activity, were analyzed at concentrations ranging from 25 to 250 µg/mL, in which the coat had inhibition variations from 12% to 40%.

The phenolic compounds present in vegetables show a high antioxidant activity [124, 125]. Literature [88] evaluated the

coat phenolic extract antioxidant activity through diverse methodologies, varying the drying temperature (Table 8). These studies show that the extracts studied had a high antioxidant potential, which was compared with a BHA synthetic antioxidant.

Table 8: Evaluating the temperature variation of cashew nut coat antioxidant potential using different methodologies

Methods	Processing conditions			Control BHA ⁽¹⁾ /Catechin ⁽²⁾
	T _{ambiance}	T 70°C/6hrs	T 130°C/33min	
Lipid-oxidation TBARS (eq.MDA-malondialdehyde / kg extract)	2,75±0,34	2,34±0,31	2,75±0,18	2,12±0,15 ⁽¹⁾
Co-oxidation β-carotene / Linoleic acid (coef. Ativi. Antiox. / G extract)	370	365	345	-
Peroxidation of LDL oxidation - inhibition (%)	46,05±0,24	41,51±0,72	43,66±2,13	40,00±1,52 ⁽²⁾
Induction of DNA H ₂ O ₂ (%)	87	91	84	-
Rancimat (inflection Factor)	2,83±0,05	1,57±0,02	1,48±0,01	2,48±0,13 ⁽¹⁾

Values expressed as mean ± standard deviation (n = 3); (-) Not reviewed.

Although the coat has a high percentage of catechins in its composition as seen on Table 8 [64, 88, 96, 126], other antioxidant compounds, i.e. the tocopherol, the thiamine, and/or phenolics, may influence the quantification of the catechin, thus justifying the variations in its analysis.

The oxidation rate of lipids, proteins, and DNA by the superoxide radical is relatively low. However, its importance as an oxidative process is connected to its capacity to generate other reactive oxygen species, as hydroxyl radical (•OH), which has high reactivity.

• BIOLOGICAL ACTIVITIES

The use of plants as medicinal agents in the treatment of many diseases has been investigated since the most ancient civilizations. Many researches have been developed based on the uses of vegetables used in folk medicine, leading to new drugs. Nowadays, there is an increased global interest and demand for herbal medicines due to their extensive biological activities, while presenting less toxicity than the synthetic drugs, and lower production costs. Current estimates indicate that around 80% of people from developing countries still rely on traditional drugs obtained from many plant species [127-130].

After two centuries of studies, the natural products chemistry continues to be a challenge and an important

research field for diverse science areas (chemical, biological, medical, agronomic, botanical, and pharmacological). The motivation behind this demand resides in the high pharmacological potential observed in natural products, that can be optimized by detection, isolation, and purification processes, especially with advances in spectral techniques [infrared (IR), mass spectrometry (MS), and nuclear magnetic resonance (NMR)] to the structural elucidation of new compounds and complexes [131].

Many parts of the cashew tree (*Anacardium occidentale* L.) have been studied for their biological potential. In folk medicine, the cashew tree has been used for diverse purposes. The juice of its pseudo fruit is utilized as antipyretic and antacid, it is also used in the treatment of premature aging and in skin remineralization processes [132, 133]. The tree's root and bole have been utilized as anti-inflammatory agents and in diarrhea treatment [134]. The cashew tree leaf is widely used for diarrhea and cramps treatment. In Nigeria, cashew the tree leaf extract has been used to reduce blood pressure and blood sugar [133]. Some Surinam tribes utilize cashew seed oil as a vermicide for butterfly larvae. In Brazil, the bole shell tea is used as an astringent to stop the bleeding after teeth extraction [134]. The cashew juice and bole shell tea are common remedies for diarrhea used by local residents throughout the Amazon.

A wine made from the pseudo fruit is utilized in dysentery treatment in other Amazon areas [12, 135-137].

Hence, the therapeutic potential of the *Anacardium occidentale* L. becomes evident due to its large usage in biological activities. It has a low cost and is widely available, thus, attracting the interest of scientists aimed at taking advantage of its popular features.

Literature shows that, used in pharmacological activities, cashew tree is an anti-inflammatory plant [16, 133, 134, 138, 139], antidiabetic [140-142], inhibitor of the acetylcholinesterase enzyme [142], used in the treatment of acute gastritis and stomach ulcers [135]. Isolated, substances of the fruit are proven to inhibit tyrosinase [143]. Nonetheless, it is utilized as an antiseptic for skin diseases and vaginal astringent treatment [144-149].

All the plant parts show antimicrobial activities both to gram negative and positive bacteria, as well as a potent anti-inflammatory action when compared to acetylsalicylic acid [150]. Other plant therapeutic actions include inhibiting the formation of dental bacterial plaque, besides being utilized against *Leishmania (Viannia) brasiliensis* [12, 137, 151, 152]. Both the bole shells and leaves have a large quantity of polyphenols, especially tannins, which are primarily responsible for the pharmacological properties [146, 153, 154].

Anacardic acids- phytochemical components of cashew nut extract- present in all parts (shell, coat, and nut), have been used to reduce cellular aging effect and helps to inhibit and kill some cancer cells [155, 156]. Besides the anacardic acids, these tests included the cardol, hydroxybenzoic acid, salicylic acid, and kaempferol tannins.

• ANTIMICROBIAL ACTIVITY

The indiscriminate use of antimicrobials, commonly marketed and utilized in infectious disease treatments, and the microorganism mutation observed in recent years, led to the increase of microbial resistance to multiple drugs [157, 158]. Secondary metabolites and complex structure compounds, such as alkaloids, terpenoids, and phenolics, as

well as their derivatives, have been under investigation to confirm their medicinal and healing properties.

Studies show that different parts of *Anacardium occidentale* L. may present activities against some bacteria and fungi. The activity of the *Anacardium occidentale* L. was observed *in vitro* [159]. The shell extract on Streptococcus species, presented activities for the *S. mitis*, *S. mutans*, and *S. sanguis*, present in the gingival supra bacterial biofilm. The extract may be used therapeutically in the odontology as an antibacterial agent. The antifungal potential of the cashew shell extracts against *C. tropicalis* and *C. stellatoidea* was observed [160].

The antibacterial activity of the cashew nut extract using 10 bacterial lineages was evaluated [161]. The researchers observed that 4 bacteria (*Proteus mirabilis*, *Shigella sonnei*, *Staphylococcus aureus*, and *Staphylococcus spp. Coagulase*) were sensitive to the extract, and the others (*Escherichia coli*, *Enterobacter aerogenes*, *Streptococcus pyogenes*, *Klebsiella pneumoniae*, *Providencia spp.*, and *Pseudomonas aeruginosa*) were resistant to cashew nut extract.

Studies performed with hydroalcoholic extract coming from the cashew tree (*Anacardium occidentale* Linn.) produced significant antimicrobial activity *in vitro* on the *Staphylococcus aureus* lineages from hospital human rise resistant and sensitive to methicillin, becoming an effective therapeutic alternative for infections caused by *Staphylococcus aureus*. This low cost alternative is easily accessible to the population, as it's use is already widespread in folk medicine [162]. The extract of cashew tree leaves observed showed activity against *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Bacillus cereus*, *Salmonella enterica serovar typhimurium*, and *Klebsiella pneumoniae*, demonstrating the high efficiency of the plant [163].

Polyphenols isolated from the nut, like the anacardio acid and the cardanol, showed high antimicrobial inhibition in fungi and bacteria (*C. albicans*, *C. Utilis*, *S. aureus*, and *S. mutans*) [164, 165]. Phenolic acids such as caffeic, ferulic and p-coumaric acid have been reported as antifungal [166,

167]. Studies [168] evaluated coat extracts (acetone and ethanol) and verified that the coat offers antimicrobial activity against some bacteria considered pathogenic to humans, being two gram-positive bacteria (*Micrococcus luteus* and *Staphylococcus aureus*) and four gram-negative

bacteria (*Salmonella typhi*, *Klebsiella pneumoniae*, *Escherichia coli*, and *Pseudomonas aeruginosa*). Table 9 shows the results comparing the coat extract with the commercial antibiotic.

Table 9: Comparison of the inhibition zones by coat extracts in comparison to a commercially traded antibiotic

Microorganism	Extract (6mg/100µL)			Amicacina (30 mcg / mL)
	Aquoso	Acetona	Etanólico	
<i>Escherichia coli</i>	13	21	34	27
<i>Klebsiella pneumoniae</i>	-	24	18	29
<i>Micrococcus luteus</i>	14	24	28	24
<i>Pseudomonas aeruginosa</i>	-	19	31	29
<i>Salmonella typhi</i>	-	22	27	24
<i>Staphylococcus aureus</i>	11	21	27	34

(-) Not analysed

Source: [168].

The results of Table 9 show that the three extracts (water, acetone, and ethanol) of *Anacardium occidentale* exhibited, effectively, antimicrobial activity against six bacterial strains with zones of inhibition that ranged from 12.0 to 34.0mm. The extracts showed a wide activity spectrum compared with a pattern antibiotic, Amikacin (30 mcg/mL) and the coat ethanolic extract obtained a higher potential in comparison with other extracts and with the Amikacin.

• CHEMICAL APPLICATIONS

The cashew nut coat has diverse constituents with applications covering the food industry, medicinal, petrochemical, cosmetics, and other areas of the chemical industry.

In the coat chemical composition it has a significant amount of fatty acids (stearic, oleic, and linoleic acid), which are responsible for many applications. Plant extracts, which have stearic acid in their compositions are commonly used in processes that are intended to form a stable and resistant base, like the cosmetics segment, the production of vegetable creams and chewing gums, and the textile industry [169-171].

Vegetable oils rich in oleic acid were studied for the possible inhibition and/or reduction of mammary tumors and on the cervix, these were induced in rats and showed significant and promising results for future work [172-187].

There are many applications for phenolic compounds. Besides phenol and other common metabolic compounds, many derivatives with diverse applications, are available, among them are: coadjuvant function at inflammatory processes treatments [188]; preservative in foods; hypoglycemic activity; and inhibitory effect on enzymatic activity and on platelet aggregation [188], blockage in the formation of cells responsible for diseases such as cancer, atherosclerosis, arthritis, diabetes, cardiovascular diseases, and processes responsible for the body aging; and antioxidants, among others [110, 189-195]. The antioxidant activity is usually attributed to soluble phenolic compounds of small chains. Researchs suggests that polyphenols with high molecular weight, such as prothocyanidins and hydrolysable tannins, are 15 to 30 times more effective than simple phenols [195].

Studies involving catechins have demonstrated its high antioxidant potential [196-202] besides the antimicrobial

inhibition [203-207], antimutagenic [208, 209], melanoma and metastasis inhibition [210-213], in the treatment of diseases like hepatitis [214], HIV [215-217], hypertension [218], and obesity [219, 220], respiratory tract [221]. Due to their antioxidant potential mentioned, the catechins have extended applications in both human health as in food applications and industry to the stability of oils and fats, acting against thermal and oxidative stress [222-227].

Besides the biological applications in antioxidant and antimicrobial activities, cashew nut coat extracts, rich in tannins, were studied [34], in phenol-formaldehyde resin formulations for the varnishes preparation. The resins were uniform, with smooth coverage without pores or any other defects, thus, providing a good quality varnish (enamels with good brightness, flexibility, scratch, and corrosion resistance).

• FINAL CONSIDERATIONS

This review reported the potential applications of the cashew nut coat. Despite being a waste of the cashew nut processing, due to its diverse chemical constituents it presents potential benefits to human health and other relevant economical possibilities. This chemical potential has been reported repeatedly in the literature when addressing the use of important fatty acids, phenolic, tannins, catechins, and bioactive compounds (β -carotene, lutein, α -zeaxanthin, α -tocopherol, γ -tocopherol, and thiamine). Its compounds are responsible for antioxidant, antimicrobial and biological activities, already proven in the literature. Thus, the cashew nut coat has diverse perspective for application in many areas of industry as pharmaceuticals, cosmetics, petroleum or food.

5. REFERENCES

1. Lubi, M.C. and E.T. Thachil, *Cashew nut shell liquid (CNSL)-a versatile monomer for polymer synthesis. Designed Monomers and Polymers*, 2000. 3(2): p. 123-153.
2. Paramashivappa, R., et al., *Novel method for isolation of major phenolic constituents*

from cashew (Anacardium occidentale L.) Nut shell liquid. Journal of Agricultural and Food Chemistry, 2001. 49(5): p. 2548-2551.

3. Das, P., T. Sreelatha, and A. Ganesh, *Bio oil from pyrolysis of cashew nut shell-characterisation and related properties. Biomass and Bioenergy*, 2004. 27(3): p. 265-275.
4. Mazzetto, S.E., D. Lomonaco, and G. Mele, *Óleo da castanha de caju: oportunidades e desafios no contexto do desenvolvimento e sustentabilidade industrial. Química Nova*, 2009. 32(3): p. 732-741.
5. CODEVASF, 2012.
6. IBGE, *Instituto Brasileiro de Geografia e Estatística (IBGE)*. 2011.
7. Pawar, S. and S. Pal, *Analgesic and anti-inflammatory activity of Anacardium occidentale Root extracts. Hamdard Medicus (Pakistan)*, 2002.
8. Yusuf, S., M. Aliyu, and R. Ndanosa, *Effect of aqueous extract of Anacardium occidentale (L) stem bark on sodium and chloride transport in the rabbit colon. J. Med. Plant. Res*, 2009. 3(6): p. 493-497.
9. Pell, S.K., *Molecular systematics of the cashew family (Anacardiaceae)*. 2004, Faculty of the Louisiana State University and Agricultural and Mechanical College in partial fulfillment of the requirements for the degree of Doctor of Philosophy in The Department of Biological Sciences by Susan Katherine Pell BS, St. Andrews Presbyterian College.

10. Florêncio, G.V.S.F.M., et al., *O polissacarídeo do Anacardium occidentale L. na fase inflamatória do processo cicatricial de lesões cutâneas*. *Ciência Rural*, 2006. **36**(1).
11. Sokeng, S., et al., *Hypoglycemic effect of Anacardium occidentale L. methanol extract and fractions on streptozotocin-induced diabetic rats*. *Research journal of medicine and medical sciences*, 2007. **2**(2): p. 133-137.
12. Akinpelu, D.A., *Antimicrobial activity of Anacardium occidentale bark*. *Fitoterapia*, 2001. **72**(3): p. 286-287.
13. Gonçalves, G.M.S. and J. Gobbo, *Antimicrobial effect of anacardium occidentale extract and cosmetic formulation development*. *Brazilian Archives of Biology and Technology*, 2012. **55**(6): p. 843-850.
14. Melo Cavalcante, A.A., et al., *Mutagenicity, antioxidant potential, and antimutagenic activity against hydrogen peroxide of cashew (Anacardium occidentale) apple juice and cajuina*. *Environmental and Molecular Mutagenesis*, 2003. **41**(5): p. 360-369.
15. Trevisan, M.T.S., et al., *Characterization of alkyl phenols in cashew (Anacardium occidentale) products and assay of their antioxidant capacity*. *Food and Chemical Toxicology*, 2006. **44**(2): p. 188-197.
16. Olajide, O.A., et al., *Effects of Anacardium occidentale stem bark extract on in vivo inflammatory models*. *Journal of Ethnopharmacology*, 2004. **95**(2-3): p. 139-142.
17. Vanderlinde, F.A., et al., *Evaluation of the antinociceptive and anti-inflammatory effects of the acetone extract from Anacardium occidentale L.* *Brazilian Journal of Pharmaceutical Sciences*, 2009. **45**(3): p. 437-442.
18. Konan, N.A., et al., *Acute, subacute toxicity and genotoxic effect of a hydroethanolic extract of the cashew (Anacardium occidentale L.)*. *Journal of Ethnopharmacology*, 2007. **110**(1): p. 30-38.
19. Ushanandini, S., et al., *The anti-ophidian properties of Anacardium occidentale bark extract*. *Immunopharmacology and Immunotoxicology*, 2009. **31**(4): p. 607-615.
20. Dahake, A.P., V.D. Joshi, and A.B. Joshi, *Antimicrobial screening of different extract of Anacardium occidentale Linn. leaves*. *Int J ChemTech Res*, 2009. **1**: p. 856-858.
21. Sankara Subramanian, S., K.J. Joseph, and A.G.R. Nair, *Polyphenols of anacardium occidentale*. *Phytochemistry*, 1969. **8**(3): p. 673.
22. Chaves, M.H., et al., *Total phenolics, antioxidant activity and chemical constituents from extracts of Anacardium occidentale L., Anacardiaceae*. *Revista Brasileira de Farmacognosia*, 2010. **20**(1): p. 106-112.
23. Murthy, S.S.N., A.S.R. Anjaneyulu, and L. Ramachandra Row, *Chemical examination*

- of *Anacardium occidentale*. Isolation and structure determination of a novel biflavonoid-C-glycoside. *Planta Medica*, 1982. **45**(1): p. 3-10.
24. Murthy, S.S.N., *Semecarpuf flavanone-A new biflavanone from Semecarpus anacardium* Linn. *Proceedings of the Indian Academy of Sciences - Chemical Sciences*, 1986. **97**(1): p. 63-69.
25. Rahman, W., et al., *Prunin-6"-O-p-coumarate, a new acylated flavanone glycoside from Anacardium occidentale*. *Phytochemistry*, 1978. **17**(6): p. 1064-1065.
26. Kubo, I., et al., *Antioxidant activity of anacardic acids*. *Food Chemistry*, 2006. **99**(3): p. 555-562.
27. Kamath, V. and P.S. Rajini, *The efficacy of cashew nut (Anacardium occidentale L.) skin extract as a free radical scavenger*. *Food Chemistry*, 2007. **103**(2): p. 428-433.
28. Paiva, F.d.A., D.d.S. Garrutti, and R. da SILVA NETO, *Aproveitamento industrial do caju*. *Embrapa Agroindústria Tropical. Documentos*, 2000.
29. Lima, C.S.M., et al., *Chemical characteristics of cape-gooseberry fruits in different sepal colors and training systems*. *Revista Brasileira de Fruticultura*, 2009. **31**(4): p. 1060-1068.
30. Neto, A.B.T., et al., *Kinetic and physico-chemical characterization of cashew (Anacardium occidentale L.) wine*. *Quimica Nova*, 2006. **29**(3): p. 489-492.
31. Patel, R.N., S. Bandyopadhyay, and A. Ganesh, *Extraction of cashew (Anacardium occidentale) nut shell liquid using supercritical carbon dioxide*. *Bioresource Technology*, 2006. **97**(6): p. 847-853.
32. Andrade, T.D.J.A.D.S., et al., *Antioxidant properties and chemical composition of technical Cashew Nut Shell Liquid (tCNSL)*. *Food Chemistry*, 2011. **126**(3): p. 1044-1048.
33. Hurtado, I., *Poisonous Anacardiaceae of South America*. *Clinics in Dermatology*, 1986. **4**(2): p. 183-190.
34. Kumar, K.P.V. and M.G. Sethuraman, *Studies on oleoresinous varnishes and their natural precursors*. *Progress in Organic Coatings*, 2004. **49**(3): p. 244-251.
35. Mohod, A.G., Y.P. Khandetod, and A.G. Powar, *Processed cashew shell waste as fuel supplement for heat generation*. *Energy for Sustainable Development*, 2008. **12**(4): p. 73-76.
36. Câmara, C.I., et al., *Quantitative analysis of boldine alkaloid in natural extracts by cyclic voltammetry at a liquid-liquid interface and validation of the method by comparison with high performance liquid chromatography*. *Talanta*, 2010. **83**(2): p. 623-630.
37. Akinnusi, F., et al., *Carcass characteristics and Sensory Evaluation of Meat from Rabbits fed Cashew-nut residue based diets*. *ASSET: An International Journal (Series A)*, 2010. **7**(1): p. 19-25.
38. Gualberto Filho, A., Prof^o. Francisca Jeanne Sidrim de Figueiredo.

39. Lima, J.R., D.S. Garruti, and L.M. Bruno, *Physicochemical, microbiological and sensory characteristics of cashew nut butter made from different kernel grades-quality. LWT - Food Science and Technology*, 2012. **45**(2): p. 180-185.
40. Russell, R.W., D.M. Warburton, and D.S. Segal, *Behavioral tolerance during chronic changes in the cholinergic system. Commun. Behav. Biol*, 1969. **4**: p. 121-128.
41. Cabral, T.d.M., *Avaliação dos constituintes e do potencial mutagênico do material particulado oriundo do beneficiamento artesanal da castanha do caju*. 2010, Universidade de São Paulo.
42. Jain, R.K. and S. Kumar, *Development of a cashew nut sheller. Journal of Food Engineering*, 1997. **32**(3): p. 339-345.
43. de Assis Paiva, F.F. and R.M. da Silva Neto, *Processamento industrial da castanha-de-caju*.
44. Lima, S.d.S., et al., *Nível tecnológico e fatores de decisão para adoção de tecnologia na produção de caju no Ceará*. 2010.
45. INAMASU, R., C. BISCEGLI, and F.d.A. PAIVA, *Máquina pneumática para abrir castanha-de-cajú. Embrapa Instrumentação Agropecuária. Comunicado Técnico*, 2006.
46. Azam-Ali, S. and E. Judge, *Small-scale cashew nut processing. Coventry (UK): ITDG Schumacher Centre for Technology and Development Bourton on Dunsmore*, 2001.
47. de Oliveira, V.H., *Nutrição mineral do cajueiro*. 1995: EMBRAPA-CNPAT.
48. Bishop, G.J., *Refining the plant steroid hormone biosynthesis pathway. Trends in plant science*, 2007. **12**(9): p. 377-380.
49. Moraes, F.P., *ALIMENTOS FUNCIONAIS E NUTRACÊUTICOS: DEFINIÇÕES, LEGISLAÇÃO E BENEFÍCIOS À SAÚDE. Revista Eletrônica de Farmácia*, 2007. **3**(2).
50. Grangeia, C., et al., *Effects of trophism on nutritional and nutraceutical potential of wild edible mushrooms. Food Research International*, 2011. **44**(4): p. 1029-1035.
51. Laughton, M.J., et al., *Inhibition of mammalian 5-lipoxygenase and cyclooxygenase by flavonoids and phenolic dietary additives: relationship to antioxidant activity and to iron ion-reducing ability. Biochemical pharmacology*, 1991. **42**(9): p. 1673-1681.
52. Quettier-Deleu, C., et al., *Phenolic compounds and antioxidant activities of buckwheat (*Fagopyrum esculentum* Moench) hulls and flour. Journal of ethnopharmacology*, 2000. **72**(1): p. 35-42.
53. Sadik, C.D., H. Sies, and T. Schewe, *Inhibition of 15-lipoxygenases by flavonoids: structure-activity relations and mode of action. Biochemical Pharmacology*, 2003. **65**(5): p. 773-781.
54. Andreescu, S. and O.A. Sadik, *Correlation of analyte structures with biosensor responses using the detection of phenolic*

- estrogens as a model. *Analytical chemistry*, 2004. **76**(3): p. 552-560.
55. Ashidate, K., et al., Gentisic acid, an aspirin metabolite, inhibits oxidation of low-density lipoprotein and the formation of cholesterol ester hydroperoxides in human plasma. *European journal of pharmacology*, 2005. **513**(3): p. 173-179.
56. Ivanova, D., et al., Polyphenols and antioxidant capacity of Bulgarian medicinal plants. *Journal of Ethnopharmacology*, 2005. **96**(1): p. 145-150.
57. Kwon, Y.-I., et al., Inhibition of *Staphylococcus aureus* by phenolic phytochemicals of selected clonal herbs species of Lamiaceae family and likely mode of action through proline oxidation. *Food Biotechnology*, 2007. **21**(1): p. 71-89.
58. Wojdyło, A., J. Oszmiański, and R. Czemerys, Antioxidant activity and phenolic compounds in 32 selected herbs. *Food chemistry*, 2007. **105**(3): p. 940-949.
59. M Calderon-Montano, J., et al., A review on the dietary flavonoid kaempferol. *Mini reviews in medicinal chemistry*, 2011. **11**(4): p. 298-344.
60. Donkoh, A., et al., Evaluation of nutritional quality of dried cashew nut testa using laboratory rat as a model for pigs. *The Scientific World Journal*, 2012. **2012**.
61. King, A. and G. Young, Characteristics and occurrence of phenolic phytochemicals. *Journal of the American Dietetic Association*, 1999. **99**(2): p. 213-218.
62. Taiz, L. and E. Zeiger, *Plant Physiology*. 2006. 672.
63. Simões, C., et al., In review in Portuguese, *Farmacognosia: Da planta ao medicamento* 5th edn. Universidade/UFRGS/Edda/EFSC, Rio Grande do Sul, Brazil, 2007: p. 49-108.
64. Trox, J., et al., Bioactive compounds in cashew nut (*anacardium occidentale* L.) kernels: Effect of different shelling methods. *Journal of Agricultural and Food Chemistry*, 2010. **58**(9): p. 5341-5346.
65. Morais, S., et al., Highly unsaturated fatty acid synthesis in Atlantic salmon: Characterization of ELOVL5- and ELOVL2-like elongases. *Marine Biotechnology*, 2009. **11**(5): p. 627-639.
66. Tanamati, A., et al., Comparative study of total lipids in beef using chlorinated solvent and low-toxicity solvent methods. *Journal of the American Oil Chemists' Society*, 2005. **82**(6): p. 393-397.
67. Maia, G. and J. Stull, Fatty acid and lipid composition of cashews (*Anacardium occidentale* L.). *Ciencia Agronomica*, 1977. **7**: p. 49-51.
68. Trox, J., et al., Catechin and epicatechin in testa and their association with bioactive compounds in kernels of cashew nut (*Anacardium occidentale* L.). *Food Chemistry*, 2011. **128**(4): p. 1094-1099.
69. Thurnhofer, S., K. Lehnert, and W. Vetter, Exclusive quantification of methyl-branched fatty acids and minor 18: 1-

- isomers in foodstuff by GC/MS in the SIM mode using 10, 11-dichloroundecanoic acid and fatty acid ethyl esters as internal standards. *European Food Research and Technology*, 2008. **226**(5): p. 975-983.
70. Scherz, H. and F. Senger, *Food composition and nutrition tables*. 1994: Medpharm GmbH Scientific Publishers.
71. Boye, J.I., L. L'Hocine, and S.H. Rajamohamed, *Processing foods without soybean ingredients. Allergen management in the food industry*, 2010: p. 355-391.
72. Gunstone, F.D., *5 Minor Specialty Oils. Nutraceutical and Specialty Lipids and their Co-Products*, 2006: p. 91.
73. Kota, L., *Total folate in peanuts and peanut products*. 2008, University of Georgia.
74. Benatti, P., et al., *Polyunsaturated fatty acids: biochemical, nutritional and epigenetic properties*. *Journal of the American College of Nutrition*, 2004. **23**(4): p. 281-302.
75. Dixon, R.A., *Plant natural products: the molecular genetic basis of biosynthetic diversity*. *Current opinion in biotechnology*, 1999. **10**(2): p. 192-197.
76. Peres, L., *Metabolismo secundário*. Disponível no site, 2004.
77. Naczki, M. and F. Shahidi, *Extraction and analysis of phenolics in food*. *Journal of Chromatography A*, 2004. **1054**(1-2): p. 95-111.
78. Matos, F.d.A., *Introdução à fitoquímica experimental*. 1997: edições UFC.
79. Kannan, V.R., et al., *Elementary chemical profiling and antifungal properties of cashew (Anacardium occidentale L.) Nuts*. *Botany Research International*, 2009. **2**(4): p. 253-257.
80. Tedong, L., et al., *Antihyperglycemic and renal protective activities of Anacardium occidentale (Anacardiaceae) leaves in streptozotocin induced diabetic rats*. *African Journal of Traditional, Complementary and Alternative Medicines*, 2006. **3**(1): p. 23-35.
81. Benavente-Garcia, O., et al., *Antioxidant activity of phenolics extracted from Olea europaea L. leaves*. *Food Chemistry*, 2000. **68**(4): p. 457-462.
82. Soong, Y.Y. and P.J. Barlow, *Antioxidant activity and phenolic content of selected fruit seeds*. *Food Chemistry*, 2004. **88**(3): p. 411-417.
83. Rockenbach, I.I., et al., *Influência do solvente no conteúdo total de polifenóis, antocianinas e atividade antioxidante de extratos de bagaço de uva (Vitis vinifera) variedades Tannat e Ancelota*. *Ciência e Tecnologia de Alimentos*, 2008. **28**: p. 238-244.
84. Chandrasekara, N. and F. Shahidi, *Antioxidative potential of cashew phenolics in food and biological model systems as affected by roasting*. *Food Chemistry*, 2011. **129**(4): p. 1388-1396.
85. Hayase, F., et al., *Scavenging of active oxygens by melanoidins*. *Agricultural and Biological Chemistry*, 1989. **53**(12): p. 3383-3385.

86. Jeong, S.M., et al., *Effect of seed roasting conditions on the antioxidant activity of defatted sesame meal extracts*. *Journal of food science*, 2004. **69**(5): p. C377-C381.
87. Şahin, H., et al., *Effect of roasting process on phenolic, antioxidant and browning properties of carob powder*. *European Food Research and Technology*, 2009. **230**(1): p. 155-161.
88. Chandrasekara, N. and F. Shahidi, *Effect of roasting on phenolic content and antioxidant activities of whole cashew nuts, kernels, and testa*. *Journal of Agricultural and Food Chemistry*, 2011. **59**(9): p. 5006-5014.
89. Shahidi, F., C. Alasalvar, and C.M. Liyana-Pathirana, *Antioxidant phytochemicals in hazelnut kernel (Corylus avellana L) and hazelnut byproducts*. *Journal of Agricultural and Food Chemistry*, 2007. **55**(4): p. 1212-1220.
90. Yu, J., et al., *Peanut skin procyanidins: Composition and antioxidant activities as affected by processing*. *Journal of Food Composition and Analysis*, 2006. **19**(4): p. 364-371.
91. Locatelli, M., et al., *Total antioxidant activity of hazelnut skin (Nocciola Piemonte PGI): Impact of different roasting conditions*. *Food Chemistry*, 2010. **119**(4): p. 1647-1655.
92. Oldoni, T.L.C., *Isolamento e identificação de compostos com atividade antioxidante de uma nova variedade de própolis brasileira produzida por abelhas da espécie Apis mellifera*. 2007, *Escola Superior de Agricultura "Luiz de Queiroz"*.
93. Melo, P.S., et al., *Phenolic composition and antioxidant activity of agroindustrial residues*. *Ciência Rural*, 2011. **41**(6): p. 1088-1093.
94. Colaric, M., et al., *Phenolic acids, syringaldehyde, and juglone in fruits of different cultivars of Juglans regia L*. *Journal of Agricultural and Food Chemistry*, 2005. **53**(16): p. 6390-6396.
95. Wijeratne, S.S.K., M.M. Abou-Zaid, and F. Shahidi, *Antioxidant polyphenols in almond and its coproducts*. *Journal of Agricultural and Food Chemistry*, 2006. **54**(2): p. 312-318.
96. Pillai, M., K. Kedlaya, and R. Selvarangan, *Cashew seed skin as a tanning material*. *Leather Science*, 1963. **10**: p. 317.
97. Neiva, T.J., et al., *Evaluation of platelet aggregation in platelet concentrates: storage implications*. *Revista Brasileira de Hematologia e Hemoterapia*, 2003. **25**(4): p. 207-212.
98. Lakhanpal, P. and D.K. Rai, *Quercetin: a versatile flavonoid*. *Internet Journal of Medical Update*, 2007. **2**(2): p. 22-37.
99. Dubey, R.K., B.L. Mandhyan, and N.K. Khandelwal, *Steaming and pressing - and integrated approach for more oil recovery*. *Journal of the Institution of Engineers (India): Agricultural Engineering Division*, 1988. **69 pt 1**: p. 1-3.

100. Cadenas, E., *Basic mechanisms of antioxidant activity. Biofactors*, 1997. **6(4)**: p. 391-397.
101. Ferreira, I.C., et al., *Free-radical scavenging capacity and reducing power of wild edible mushrooms from northeast Portugal: Individual cap and stipe activity. Food Chemistry*, 2007. **100(4)**: p. 1511-1516.
102. Valko, M., et al., *Free radicals and antioxidants in normal physiological functions and human disease. The international journal of biochemistry & cell biology*, 2007. **39(1)**: p. 44-84.
103. Chung, K.-T., et al., *Tannins and human health: a review. Critical reviews in food science and nutrition*, 1998. **38(6)**: p. 421-464.
104. Krinsky, N.I. and E.J. Johnson, *Carotenoid actions and their relation to health and disease. Molecular aspects of medicine*, 2005. **26(6)**: p. 459-516.
105. Biesalski, H.K., *Vitamin E requirements in parenteral nutrition. Gastroenterology*, 2009. **137(5)**: p. S92-S104.
106. Awaad, A.S. and N.A. Al-Jaber, *Antioxidant natural plant. RPMP Ethnomedicine: Source & Mechanism*, 2010. **27**: p. 1-35.
107. Al-Jaber, N.A., A.S. Awaad, and J.E. Moses, *Review on some antioxidant plants growing in Arab world. Journal of Saudi Chemical Society*, 2011. **15(4)**: p. 293-307.
108. Shahidi, F., P. Janitha, and P. Wanasundara, *Phenolic antioxidants. Critical reviews in food science & nutrition*, 1992. **32(1)**: p. 67-103.
109. Mastro-Durán, R. and R. Borja-Padilla, *Antioxidant activity of natural sterols and organic acids. Grasas y Aceites*, 1993. **44(3)**: p. 208-212.
110. Brenna, O.V. and E. Pagliarini, *Multivariate analysis of antioxidant power and polyphenolic composition in red wines. Journal of Agricultural and Food Chemistry*, 2001. **49(10)**: p. 4841-4844.
111. Yildirim, A., A. Mavi, and A.A. Kara, *Determination of antioxidant and antimicrobial activities of Rumex crispus L. extracts. Journal of agricultural and food chemistry*, 2001. **49(8)**: p. 4083-4089.
112. Zamora, R., M.M. León, and F.J. Hidalgo, *Free radical-scavenging activity of nonenzymatically-browned phospholipids produced in the reaction between phosphatidylethanolamine and ribose in hydrophobic media. Food Chemistry*, 2011. **124(4)**: p. 1490-1495.
113. Brand-Williams, W., M. Cuvelier, and C. Berset, *Use of a free radical method to evaluate antioxidant activity. LWT-Food Science and Technology*, 1995. **28(1)**: p. 25-30.
114. Alam, M.N., N.J. Bristi, and M. Rafiqzaman, *Review on in vivo and in vitro methods evaluation of antioxidant activity. Saudi Pharmaceutical Journal*, 2013. **21(2)**: p. 143-152.
115. Badarinath, A., et al., *A review on in-vitro antioxidant methods: comparisons, correlations and considerations.*

- International Journal of PharmTech Research*, 2010. **2**(2): p. 1276-1285.
116. Sánchez de Medina , V.n., et al., *Quality and stability of edible oils enriched with hydrophilic antioxidants from the olive tree: The role of enrichment extracts and lipid composition. Journal of agricultural and food chemistry*, 2011. **59**(21): p. 11432-11441.
117. Antolovich, M., et al., *Methods for testing antioxidant activity. Analyst*, 2002. **127**(1): p. 183-198.
118. González-Montelongo, R., M.G. Lobo, and M. González, *Antioxidant activity in banana peel extracts: testing extraction conditions and related bioactive compounds. Food Chemistry*, 2010. **119**(3): p. 1030-1039.
119. Lafka, T.-I., V. Sinanoglou, and E.S. Lazos, *On the extraction and antioxidant activity of phenolic compounds from winery wastes. Food Chemistry*, 2007. **104**(3): p. 1206-1214.
120. Li, H., et al., *Microwave-assisted extraction of phenolics with maximal antioxidant activities in tomatoes. Food Chemistry*, 2012. **130**(4): p. 928-936.
121. Zarena, A.S. and K.U. Sankar, *A study of antioxidant properties from Garcinia mangostana L. pericarp extract. Acta Sci Pol Technol Aliment*, 2009. **8**: p. 23-34.
122. Zarena, A. and K.U. Sankar, *Supercritical carbon dioxide extraction of xanthenes with antioxidant activity from Garcinia mangostana: Characterization by HPLC/LC-ESI-MS. The Journal of Supercritical Fluids*, 2009. **49**(3): p. 330-337.
123. Chaves, M.H., et al., *Fenóis totais, atividade antioxidante e constituintes químicos de extratos de Anacardium occidentale L., Anacardiaceae. Rev Bras Farmacogn*, 2010. **20**: p. 106-112.
124. Pradeep, S. and M. Guha, *Effect of processing methods on the nutraceutical and antioxidant properties of little millet (Panicum sumatrense) extracts. Food chemistry*, 2011. **126**(4): p. 1643-1647.
125. Wijeratne, S.S., M.M. Abou-Zaid, and F. Shahidi, *Antioxidant polyphenols in almond and its coproducts. Journal of Agricultural and Food Chemistry*, 2006. **54**(2): p. 312-318.
126. Chaves, M.H., et al., *Total phenolics, antioxidant activity and chemical constituents from extracts of Anacardium occidentale L., Anacardiaceae. Brazilian Journal of Pharmacognosy*, 2010. **20**(1): p. 106-112.
127. Gilani, A.H., Atta-ur-Rahman. 2005. *Trends in ethnopharmacology. J. Ethnopharmacol.* **100**: p. 43-49.
128. Atta-ur-Rahman, Z.S., et al., *Some chemical constituents of Terminalia glaucescens and their enzymes inhibition activity. Z Naturforsch*, 2005. **13**: p. 347-350.
129. Grabley, S. and R. Thiericke, *Bioactive agents from natural sources: trends in discovery and application, in Thermal Biosensors, Bioactivity, Bioaffinity*. 1999, Springer. p. 101-154.

130. Grabley, S. and R. Thiericke, *The impact of natural products on drug discovery. Drug discovery from nature. Springer, New York Berlin Heidelberg, 1999: p. 3-37.*
131. Desai, N.C., et al., *Degumming of vegetable oil by membrane technology. Indian Journal of Chemical Technology, 2002. 9(6): p. 529-534.*
132. Patro, C. and R. Behera, *Cashew helps to fix sand dunes in Orissa. Indian Farming, 1979. 28(12): p. 31-32.*
133. Esimone, C., J. Okonta, and C. Ezugwu, *Blood sugar lowering effect of Anacardium occidentale leaf extract in experimental rabbit model. Journal of Natural Remedies, 2001. 1(1): p. 60-63.*
134. Thomas, M.M.G. and J. Barbosa Filho, *Anti-inflammatory actions of tannins isolated from the bark of Anacardium occidentale L. Journal of ethnopharmacology, 1985. 13(3): p. 289-300.*
135. Kubo, J., J.R. Lee, and I. Kubo, *Anti-Helicobacter pylori agents from the cashew apple. Journal of Agricultural and Food Chemistry, 1999. 47(2): p. 533-537.*
136. Laurens, A., S. Mboup, and P. Giono-Barber, *Study of antimicrobial activity of Anacardium occidentale L. Annales Pharmaceutiques Francaises, 1982. 40(2): p. 143-146.*
137. Kudi, A.C., et al., *Screening of some Nigerian medicinal plants for antibacterial activity. Journal of Ethnopharmacology, 1999. 67(2): p. 225-228.*
138. Falcão, H.d.S., et al., *Review of the plants with anti-inflammatory activity studied in Brazil. Revista Brasileira de Farmacognosia, 2005. 15(4): p. 381-391.*
139. Thomas, M.L.R.M.G. and J.M.B. Filho, *Anti-inflammatory actions of tannins isolated from the bark of Anacardium occidentale L. Journal of Ethnopharmacology, 1985. 13(3): p. 289-300.*
140. Oliveira, F.d. and M.L. Saito, *Alguns vegetais brasileiros empregados no tratamento da diabetes. Rev. bras. farmacogn, 1989. 2: p. 170-96.*
141. Kamtchouing, P., et al., *Protective role of Anacardium occidentale extract against streptozotocin-induced diabetes in rats. Journal of ethnopharmacology, 1998. 62(2): p. 95-99.*
142. Barbosa-Filho, J.M., et al., *Plants and their active constituents from South, Central, and North America with hypoglycemic activity. Revista Brasileira de Farmacognosia, 2005. 15(4): p. 392-413.*
143. Kubo, I., I. Kinst-Hori, and Y. Yokokawa, *Tyrosinase inhibitors from Anacardium occidentale fruits. Journal of Natural Products, 1994. 57(4): p. 545-551.*
144. AGUIAR, F. and L. Lins, *Ação hipoglicemiante da entrecasca de Anacardium occidentale L. An. Fac. Med. Univ. Recife, 1958. 18(1): p. 263.*
145. Aguiar, F., J. Cardoso, and R. Azoubel, *Novas considerações sobre o efeito hipoglicemiante da Anacardium*

- occidentale* L. *An. Fac. Med. Univ. Recife*, 1959. **19**(1): p. 353.
146. Diniz, M., et al., *Memento fitoterápico. As plantas como alternativa Terapêutica: aspectos populares e científicos*. João Pessoa: Editora Universitária/UFPB, 1998.
147. Barrett, B., *Medicinal plants of Nicaragua's Atlantic coast*. *Economic Botany*, 1994. **48**(1): p. 8-20.
148. Okamoto, M.K.H., *Estudo das atividades cicatrizante e antimicrobiana do extrato glicólico e do gel de Psidium guajava L. e estudo da estabilidade do gel*. 2010, Universidade de São Paulo.
149. de Melo, A.F.M., et al., *Avaliação da toxicidade subcrônica do extrato bruto seco de Anacardium occidentale Linn em cães-DOI: 10.4025/actascihealthsci.v28i1.1112*. *Acta Scientiarum. Health Science*, 2008. **28**(1): p. 37-41.
150. MAIA, G.C., *Aproveitamento industrial do caju (Anacardium occidentale); relatório final*. 1982: NUTEC.
151. Lim, T., *Anacardium occidentale*, in *Edible Medicinal and Non-Medicinal Plants*. 2012, Springer. p. 45-68.
152. Araujo, J.R.G., et al., *Imbibition and position of seed on the germination of seedlings of dwarf-precocious cashew rootstocks*. *Revista Brasileira de Fruticultura*, 2009. **31**(2): p. 552-558.
153. Haslam, J. and H.A. Willis, *Identification and analysis of plastics*, in *Identification and analysis of plastics*. 1965, Van Nostrand. p. 483.
154. Mota, M., *Estudos antiinflamatório e análise química da casca do Anacardium occidentale L. Estudo antiinflamatório e análise química da casca do Anacardium occidentale L*, 1982.
155. de SOUZA, C.P., et al., *O USO DA CASCA DA CASTANHA DO CAJU, Anacardium occidentale, COMO MOLUSCICIDA ALTERNATIVO*. *Revista do Instituto de Medicina Tropical de São Paulo*, 1992. **34**(5): p. 459-466.
156. Mendes, N.M., et al., *Atividade moluscicida da mistura de ácidos 6-n-alquil salicílicos (ácido anacárdico) e dos seus complexos com cobre (II) e chumbo (II)*. *Rev. Soc. bras. Med. trop*, 1990. **23**: p. 217-223.
157. Amorozo, M.C.d.M., *Use and diversity of medicinal plants in Santo Antonio do Leverger, MT, Brazil*. *Acta Botanica Brasílica*, 2002. **16**(2): p. 189-203.
158. Nascimento, G.G., et al., *Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria*. *Brazilian journal of microbiology*, 2000. **31**(4): p. 247-256.
159. Pereira, J.V., et al., *In vitro antimicrobial activity of an extract from Anacardium occidentale Linn. on Streptococcus mitis, Streptococcus mutans and Streptococcus sanguis*. *Odontol. clín.-cient*, 2006. **5**(2): p. 137-141.
160. Araújo, C.R.F., et al., *Atividade antifúngica in vitro da casca do anacardium occidentale linn. sobre*

- leveduras do gênero *Candida*. *Arq. odontol*, 2005. **41**(03): p. 263-270.
161. Gonçalves, J.L.S., et al., *In vitro* anti-rotavirus activity of some medicinal plants used in Brazil against diarrhea. *Journal of Ethnopharmacology*, 2005. **99**(3): p. 403-407.
162. Silva, A.M.M., et al., Alkaloids from *Prosopis juliflora* leaves induce glial activation, cytotoxicity and stimulate NO production. *Toxicol*, 2007. **49**(5): p. 601-614.
163. Braga, F.G., et al., Antileishmanial and antifungal activity of plants used in traditional medicine in Brazil. *Journal of Ethnopharmacology*, 2007. **111**(2): p. 396-402.
164. Kubo, I., et al., Antitumor agents from the cashew (*Anacardium occidentale*) apple juice. *Journal of Agricultural and Food Chemistry*, 1993. **41**(6): p. 1112-1115.
165. LIMA, C.A.d.A., G.M. Pastore, and E.D.P.d.A. LIMA, Estudo da atividade antimicrobiana dos ácidos anacárdicos do óleo da casca da castanha de caju (CNSL) dos clones de cajueiro-anão-precoce CCP-76 e CCP-09 em cinco estágios de maturação sobre microrganismos da cavidade bucal. *Sociedade Brasileira de Ciência e Tecnologia de Alimentos*, 2000.
166. Dragland, S., et al., Several culinary and medicinal herbs are important sources of dietary antioxidants. *The Journal of nutrition*, 2003. **133**(5): p. 1286-1290.
167. Moure, A., et al., Natural antioxidants from residual sources. *Food chemistry*, 2001. **72**(2): p. 145-171.
168. Doss, V.A. and K.P. Thangavel, ANTIOXIDANT AND ANTIMICROBIAL ACTIVITY USING DIFFERENT EXTRACTS OF *ANACARDIUM OCCIDENTALE* L. 2011.
169. DA CASTANHA-DE-CAJU, A.A., A Amêndoa da Castanha-de-caju: Composição e Importância dos Ácidos Graxos—Produção e Comércio Mundiais.
170. Alcântara, S.R., et al., Isotermas de adsorção do pedúnculo seco do caju. *Revista Brasileira de Engenharia Agrícola e Ambiental*, 2009. **13**(1): p. 81-87.
171. Voigt, E.L., et al., Source-sink regulation of cotyledonary reserve mobilization during cashew (*Anacardium occidentale*) seedling establishment under NaCl salinity. *Journal of Plant Physiology*, 2009. **166**(1): p. 80-89.
172. Welsch, C.W., Relationship between dietary fat and experimental mammary tumorigenesis: a review and critique. *Cancer research*, 1992. **52**(7 Supplement): p. 2040s-2048s.
173. Carroll, K. and H. Khor, Effects of level and type of dietary fat on incidence of mammary tumors induced in female Sprague-Dawley rats by 7, 12-dimethylbenz (a) anthracene. *Lipids*, 1971. **6**(6): p. 415-420.
174. Muller, W.J., et al., Single-step induction of mammary adenocarcinoma in transgenic mice bearing the activated c-

- neu oncogene. Cell, 1988. 54(1): p. 105-115.*
175. Cohen, L.A., et al., *Dietary fat and mammary cancer. I. Promoting effects of different dietary fats on N-nitrosomethylurea-induced rat mammary tumorigenesis. Journal of the National Cancer Institute, 1986. 77(1): p. 33-42.*
176. Brockman, H., H. Stack, and M. Waters, *Antimutagenicity profiles of some natural substances. Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis, 1992. 267(2): p. 157-172.*
177. Dayton, S., S. Hashimoto, and J. Wollman, *Effect of high-oleic and high-linoleic safflower oils on mammary tumors induced in rats by 7, 12-dimethylbenz (alpha) anthracene. The Journal of nutrition, 1977. 107(8): p. 1353-1360.*
178. Sundram, K., et al., *Effect of dietary palm oils on mammary carcinogenesis in female rats induced by 7, 12-dimethylbenz (a) anthracene. Cancer Research, 1989. 49(6): p. 1447-1451.*
179. Corona, G., J. Spencer, and M. Dessi, *Extra virgin olive oil phenolics: absorption, metabolism, and biological activities in the GI tract. Toxicology and industrial health, 2009. 25(4-5): p. 285-293.*
180. Escrich, E., R. Moral, and M. Solanas, *Olive oil, an essential component of the Mediterranean diet, and breast cancer. Public health nutrition, 2011. 14(12A): p. 2323-2332.*
181. Timsina, B., M. Shukla, and V.K. Nadumane, *A review of few essential oils and their anticancer property. International Journal of Shoulder Surgery, 2012. 6(2).*
182. Juan, M.E., et al., *Cancer Chemopreventive Activity of Hydroxytyrosol: A Natural Antioxidant from Olives and Olive Oil. 2010.*
183. Maggiora, M., et al., *An overview of the effect of linoleic and conjugated-linoleic acids on the growth of several human tumor cell lines. International Journal of Cancer, 2004. 112(6): p. 909-919.*
184. Welsch, C.W., et al., *Selenium and the genesis of murine mammary tumors. Carcinogenesis, 1981. 2(6): p. 519-522.*
185. Rogers, A.E. and W.C. Wetsel, *Mammary carcinogenesis in rats fed different amounts and types of fat. Cancer research, 1981. 41(9 Part 2): p. 3735-3737.*
186. Brown, R., *Effects of dietary fat on incidence of spontaneous and induced cancer in mice. Cancer research, 1981. 41(9 Part 2): p. 3741-3742.*
187. Steinmetz, K.A. and J.D. Potter, *Vegetables, fruit, and cancer prevention: a review. Journal of the American Dietetic Association, 1996. 96(10): p. 1027-1039.*
188. Maillard, M.-N. and C. Berset, *Evolution of antioxidant activity during kilning: role of insoluble bound phenolic acids of barley and malt. Journal of Agricultural and Food Chemistry, 1995. 43(7): p. 1789-1793.*

189. Yildirim, T., et al., Giant anharmonicity and nonlinear electron-phonon coupling in MgB₂: a combined first-principles calculation and neutron scattering study. *Physical review letters*, 2001. **87**(3): p. 037001.
190. Baluchnejadmojarad, T., et al., Beneficial effect of aqueous garlic extract on the vascular reactivity of streptozotocin-diabetic rats. *Journal of ethnopharmacology*, 2003. **85**(1): p. 139-144.
191. Carvalho, C.A.d., et al., Antioxidant activity of *Jacaranda decurrens* Cham., *Bignoniaceae*. *Revista Brasileira de Farmacognosia*, 2009. **19**(2B): p. 592-598.
192. Bravo, K., et al., Influence of cultivar and ripening time on bioactive compounds and antioxidant properties in Cape gooseberry (*Physalis peruviana* L.). *Journal of the Science of Food and Agriculture*, 2014.
193. Ranilla, L.G., et al., Phenolic compounds, antioxidant activity and in vitro inhibitory potential against key enzymes relevant for hyperglycemia and hypertension of commonly used medicinal plants, herbs and spices in Latin America. *Bioresource Technology*, 2010. **101**(12): p. 4676-4689.
194. Kalt, W., et al., Antioxidant capacity, vitamin C, phenolics, and anthocyanins after fresh storage of small fruits. *Journal of Agricultural and Food Chemistry*, 1999. **47**(11): p. 4638-4644.
195. Velioglu, Y., et al., Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of agricultural and food chemistry*, 1998. **46**(10): p. 4113-4117.
196. Atoui, A.K., et al., Tea and herbal infusions: their antioxidant activity and phenolic profile. *Food chemistry*, 2005. **89**(1): p. 27-36.
197. Azam, S., et al., Prooxidant property of green tea polyphenols epicatechin and epigallocatechin-3-gallate: implications for anticancer properties. *Toxicology in vitro*, 2004. **18**(5): p. 555-561.
198. Halliwell, B., et al., The characterization of antioxidants. *Food and Chemical Toxicology*, 1995. **33**(7): p. 601-617.
199. Salah, N., et al., Polyphenolic flavanols as scavengers of aqueous phase radicals and as chain-breaking antioxidants. *Archives of biochemistry and biophysics*, 1995. **322**(2): p. 339-346.
200. Tang, S., et al., Antioxidative mechanisms of tea catechins in chicken meat systems. *Food Chemistry*, 2002. **76**(1): p. 45-51.
201. Shahidi, F., et al., Endogenous antioxidants and stability of sesame oil as affected by processing and storage. *JAOCs, Journal of the American Oil Chemists' Society*, 1997. **74**(2): p. 143-148.
202. Muzolf, M., et al., pH-dependent radical scavenging capacity of green tea catechins. *Journal of agricultural and food chemistry*, 2008. **56**(3): p. 816-823.
203. Chung, K.-T., C.-I. Wei, and M.G. Johnson, Are tannins a double-edged sword in biology and health? *Trends in*

- Food Science & Technology*, 1998. **9**(4): p. 168-175.
204. Yoda, Y., et al., Different susceptibilities of *Staphylococcus* and Gram-negative rods to epigallocatechin gallate. *Journal of Infection and Chemotherapy*, 2004. **10**(1): p. 55-58.
205. Dykes, G.A., R. Amarowicz, and R.B. Pegg, Enhancement of nisin antibacterial activity by a bearberry (*Arctostaphylos uva-ursi*) leaf extract. *Food microbiology*, 2003. **20**(2): p. 211-216.
206. Yanagawa, Y., et al., A combination effect of epigallocatechin gallate, a major compound of green tea catechins, with antibiotics on *Helicobacter pylori* growth in vitro. *Current microbiology*, 2003. **47**(3): p. 0244-0249.
207. Yee, Y.K. and M.W.L. Koo, Anti-*Helicobacter pylori* activity of Chinese tea: in vitro study. *Alimentary pharmacology & therapeutics*, 2000. **14**(5): p. 635-638.
208. Gupta, S., B. Saha, and A. Giri, Comparative antimutagenic and anticlastogenic effects of green tea and black tea: a review. *Mutation Research/Reviews in Mutation Research*, 2002. **512**(1): p. 37-65.
209. Wang, Z.Y., et al., Antimutagenic activity of green tea polyphenols. *Mutation Research/Genetic Toxicology*, 1989. **223**(3): p. 273-285.
210. Kohno, M., et al., CD151 enhances cell motility and metastasis of cancer cells in the presence of focal adhesion kinase. *International journal of cancer*, 2002. **97**(3): p. 336-343.
211. Yamamoto, Y. and H. Nakamura, 1-Carboranyl-3-(2-methylaziridino)-2-propanol. Synthesis, selective uptake by B-16 melanoma, and selective cytotoxicity toward cancer cells. *Journal of medicinal chemistry*, 1993. **36**(15): p. 2232-2234.
212. Sazuka, M., et al., Inhibitory effects of green tea infusion on in vitro invasion and in vivo metastasis of mouse lung carcinoma cells. *Cancer letters*, 1995. **98**(1): p. 27-31.
213. Taniguchi, S., T. Iwamura, and T. Katsuki, Correlation between spontaneous metastatic potential and type I collagenolytic activity in a human pancreatic cancer cell line (SUIT-2) and sublines. *Clinical & experimental metastasis*, 1992. **10**(4): p. 259-266.
214. Nishida, N., et al., Amplification and overexpression of the cyclin D1 gene in aggressive human hepatocellular carcinoma. *Cancer research*, 1994. **54**(12): p. 3107-3110.
215. Nakane, H. and K. Ono, Differential inhibitory effects of some catechin derivatives on the activities of human immunodeficiency virus reverse transcriptase and cellular deoxyribonucleic and ribonucleic acid polymerases. *Biochemistry*, 1990. **29**(11): p. 2841-2845.
216. Nakane, H. and K. Ono, Differential inhibition of HIV-reverse transcriptase and various DNA and RNA polymerases

- by some catechin derivatives. in *Nucleic acids symposium series*. 1988.
217. Moore, P.S. and C. Pizza, Observations on the inhibition of HIV-1 reverse transcriptase by catechins. *Biochem. J*, 1992. **288**: p. 717-719.
218. Hara, Y. and T. Suzuki, *Administering epigallocatechin gallate, theaflavin. theaflavin mono (di) gallate*. 1989, Google Patents.
219. Rains, T.M., S. Agarwal, and K.C. Maki, *Antiobesity effects of green tea catechins: a mechanistic review. The Journal of nutritional biochemistry*, 2011. **22**(1): p. 1-7.
220. Zaveri, N.T., *Green tea and its polyphenolic catechins: medicinal uses in cancer and noncancer applications. Life sciences*, 2006. **78**(18): p. 2073-2080.
221. Ohmori, Y., et al., *Antiallergic constituents from oolong tea stem. Biological and Pharmaceutical Bulletin*, 1995. **18**(5): p. 683-686.
222. Mitsumoto, M., et al., *Addition of tea catechins and vitamin C on sensory evaluation, colour and lipid stability during chilled storage in cooked or raw beef and chicken patties. Meat Science*, 2005. **69**(4): p. 773-779.
223. O'Sullivan, A., et al., *Use of natural antioxidants to stabilize fish oil systems. Journal of Aquatic Food Product Technology*, 2005. **14**(3): p. 75-94.
224. Tang, S., et al., *Antioxidative effect of added tea catechins on susceptibility of cooked red meat, poultry and fish patties to lipid oxidation. Food Research International*, 2001. **34**(8): p. 651-657.
225. Chen, Z. and P. Chan, *Antioxidative activity of green tea catechins in canola oil. Chemistry and Physics of Lipids*, 1996. **82**(2): p. 163-172.
226. Surco-Laos, F., et al., *Influence of catechins and their methylated metabolites on lifespan and resistance to oxidative and thermal stress of Caenorhabditis elegans and epicatechin uptake. Food Research International*, 2012. **46**(2): p. 514-521.
227. González-Manzano, S., et al., *Oxidative status of stressed Caenorhabditis elegans treated with epicatechin. Journal of agricultural and food chemistry*, 2012. **60**(36): p. 8911-8916.

